

Fire Blight of Apple and Pear

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Overview

Fire blight is an important disease affecting pear and apple. Infections commonly occur during bloom or on late blooms during the three weeks following petal fall. Increased acreage of highly susceptible apple varieties on highly susceptible rootstocks has increased the danger that infected blocks will suffer significant damage. In Washington there have been minor outbreaks annually since 1991 and serious damage in about 5-10 percent of orchards in 1993, 1997, 1998, 2005, 2009, 2012, 2015, 2016, 2017 and 2018.

Casual Organism

Fire blight is caused by *Erwinia amylovora*, a gram-negative, rod-shaped bacterium. The bacterium grows by splitting its cells and this rate of division is regulated by temperature. Cell division is minimal below 50° F, and relatively slow at air temperatures between 50° to 70° F. At air temperatures above 70° F, the rate of cell division increases rapidly and is fastest at 80° F. Above 95°F cell density on and in the plant can actually decline (Pusey and Curry 2004).

Host Range

Considered a problem for apple and pear, *Erwinia amylovora* has a wide host range within Rosacea and Rubus with reports on about 200 species including crab apple, hawthorn, mountain ash and Bradford pear (Timur Momol and Aldwinckle 2000).

Signs and Symptoms

Overwintering cankers can appear black, grey or violet. Older cankers may have dry sunken tissue. If the bark is cut from the edge of an active canker, reddish flecking can be seen in the wood near the canker margin. (Teviotdale 2011).



Figure 1 Canker on apple. Photo T. DuPont, WSU.

Blossom symptoms become apparent one to two weeks after infection. The floral receptacle, ovary, and peduncles become water soaked and dull, grayish green in appearance. Later tissues shrivel and turn brown to black. During periods of high humidity, small droplets of bacterial ooze form on water-soaked and discolored tissues. Ooze droplets start creamy white, becoming amber tinted as they age (Johnson 2000).



Figure 2 Bloom symptoms 12 days after infection. Photo T. DuPont, WSU.

Shoot symptoms. Tips of shoots may wilt rapidly to form a "shepherd's crook." Leaves on diseased shoots often show blackening along the midrib and veins before becoming fully necrotic, and cling firmly to the host after death (a key diagnostic feature.) Numerous diseased shoots give a tree a burnt, blighted appearance, hence the disease name.



Figure 3 Characteristic shepherd's crook. Notice ooze. Photo T. DuPont, WSU.

Rootstocks infections usually develop near the graft union as a result of internal movement of the pathogen through the tree or from infection of root suckers. The bark of infected rootstocks may show water-soaking, purplish to black discoloration, cracking, or signs of bacterial ooze. Red-brown streaking may be apparent in cambium just under the bark. Symptoms of rootstock blight can be confused with Phytophthora collar rot. Malling 26 and 9 rootstocks are highly susceptible to fire blight (Johnson 2000).



Figure 4 Rootstock infections may appear water soaked under the bark.

Transmission and Disease Cycle

Erwinia amylovora overwinters within diseased plant tissue (e.g. cankers). In 20 to 50% of cankers active cells survive the winter (van der Zwet and Beer 1991) and when humidity is high in the spring the pathogen oozes out of these cankers. This ooze is attractive to bees, flies and other insects who transfer the blight pathogen to flowers. Pathogen cells can also be moved from old cankers to flowers by splashed and wind-blown rain. Pathogen cells multiply quickly on nutrient rich floral stigmas when temperatures are warm (70-80 F is optimal for the pathogen) (Ogawa and English 1991). Bacterial colonies can then be washed down the style into the floral cup by water (usually from rain or heavy dew) where they can invade flowers through the nectaries. Once initial blossoms are infested, insects and rain can move the pathogen to additional flowers (Pattimore et al. 2014, Johnson et al. 1993). If the pathogen is successful in infecting the developing fruit-let, the disease spreads into the cambium (just between the bark and the wood) of the tree, killing young host tissues as it progresses (Momol et al. 1998) creating characteristic strikes and cankers. Pathogen cells migrate inside the tree well ahead of visible symptoms where they can accumulate in other susceptible tissue such as one-year old shoot tips and susceptible rootstocks causing infections distant from the original infection point. *Erwinia amylovora* can also infect susceptible one and two-year-old tissue directly through wounds (e.g. insect feeding and hail) causing shoot blight infections.

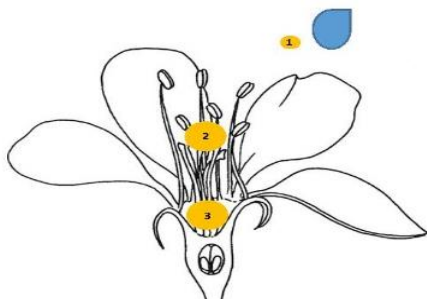


Figure 5 Pathogen cells (1) multiply on the flower stigma (2) and if rain or dew occur are washed into the floral cup (3).

Cultural Controls

Plant on resistant rootstock. Resistant rootstocks (e.g. Geneva series for apples) do not make the scion less susceptible, but will help prevent tree death from rootstock blight.

Sanitation. In winter, cut out old blight cankers as thoroughly as possible. Ideally, cut blight before you prune for tree structure so that the blighted cuttings can be removed from the orchard. Compared to cuts made in summer, winter removal cuts can be made closer to the visible canker edge. In winter the pathogen is confined to the cankered area. Cut at the next "horticulturally sensible" site below the canker. You do not need to sterilize tools when you are cutting on fully dormant trees. Late dormant copper applications may also provide orchard sanitation, reducing inocula levels going into spring (Elkins et al. 2015). During the summer, cut out blight when you see it. Make summer cuts AT LEAST 12-18" below

the edge of the visible canker. Cut more aggressively in young, vigorous trees or susceptible varieties. Removing a strike can greatly reduce further damage to the tree if cut early.

Manage the orchard environment. In addition to warm temperatures moisture is required to create infections. As little as two to three hours of wetting is sufficient to trigger infections. Manage weeds/cover crops to limit relative humidity. Do not irrigate during bloom.

Blossom removal in young blocks. Blossom removal in young blocks and removal of late blooms limits the numbers of flowers and thus reduces potential points of infection.

Keep vigor of the tree moderate. Moderating vigor will not prevent infection, but it can reduce damage to the tree.

Temperature Risk Models

The risk of fire blight infections during bloom can be calculated based on the temperature and moisture. In Washington the best prediction model is CougarBlight available at [WSU Decision Aid System for Tree Fruit \(DAS\)](http://www.wsu.edu/DecisionAidSystemforTreeFruit). This model calculates fire blight risk based on the temperature of the previous four days using the documented growth rate of the bacteria, e.g. higher risk with multiple hours above 70 F. (Pusey and Curry 2004). The model then projects risk for the next three days based on predicted temperatures. Growers can use model information to decide when to spray. If trees are likely to be blooming during an upcoming high-risk period, protective sprays are recommended (Smith and Pusey 2010).

Chemical Control Programs

There is a risk of fire blight infection any time there are flowers on the tree, the weather is warm, and wetting occurs. Watch for and protect secondary blossoms during the three weeks after petal fall, which is the most common time of fire blight infection. Most sprays only protect the blooms that are open. Protect new blooms as they open. In warm weather follow-up sprays are needed every few days.

Conventional Management

Prebloom. Fixed-copper sanitation, but only if fire blight was in the orchard last year.

Early bloom. Apply biologicals (Blossom Protect) during early bloom. If fire blight was in the orchard last year, apply two applications of the biological. Re-apply biologicals a second time if lime sulfur was applied. Lime sulfur applied during early bloom is also antimicrobial and reduces blight pressure.

Early bloom to petal fall. Watch the model. After a period of warm weather, best results are obtained when antibiotics are applied within the 24-hour window before flower wetting during a high infection risk period. Products used must contact the interior of the flowers in sufficient water and approved wetting agent to completely cover the interior. Repeated antibiotic sprays may be necessary during extended high or

extreme risk periods. One pound of any 17% oxytetracycline product per 100 gallons gives a 200-ppm solution. Kasugamycin is another effective antibiotic. Some trials have shown that a full rate of Kasugamycin and a full rate of oxytetracycline provides excellent control. Applications of less than 100 gal/A can be effective on small trees if flower interiors are well covered, but do not drop the ppm below 200 (oxytetracycline). Application by ground equipment on each row is highly recommended (aircraft is NOT recommended). Many fire blight bacteria in the Pacific Northwest are resistant to streptomycin, another registered antibiotic.

Organic Management

Prebloom. Fixed copper sanitation if fire blight was in the **orchard** last year.

Early bloom. Lime sulfur plus oil (apples only). One to two applications of biologicals (Blossom Protect). Reapply biological after lime sulfur, which is antimicrobial.

Full bloom to petal fall. Depending on the cultivar russet risk and the CougarBlight model risk follow with *Bacillus subtilis* (Serenade Opti) (most fruit safe) every 2-5 days during flower/petal fall or copper hydroxide/octanoate (e.g. Cueva, Previsto) every 2 to 6 days (less fruit safe for russet). Coppers have had higher efficacy than biologicals during bloom in Washington trials. Do not follow coppers with any products with acidifiers. Good drying conditions are important to avoid russet risk.

Petal fall to two weeks after. Continue protective programs one to two weeks post petal fall. Warm conditions during late bloom increase fire blight risk for late blooms still present.

Strategies for Improving Protective Programs

Coverage. Product efficacy is based on thorough coverage of flowers. Use tree row volume to apply appropriate volumes to cover the tree architecture in your orchard. Products applied every other row or at high speeds may have insufficient coverage and lower efficacy.

Timing. Antibiotics have the highest efficacy when applied shortly before a moisture event. Nonetheless, Kasugamycin and Streptomycin can also be applied up to 12 hours after a moisture event, but with reduced effectiveness. Streptomycin has locally systemic activity and Kasugamycin is effective on bacteria which have been washed into the floral cup but not yet invaded the flower.

pH of spray tank water. It is important to appropriately acidify spray tank water when using antibiotics (especially oxytetracycline and kasugamycin). Antibiotic efficacy reported in WSU trials is with spray tank water acidified to pH 5.6. At higher pH antibiotic degradation rate is higher and thus efficacy is often lower. For example, in one trial Kasugamycin

reduced bacteria by 86 to 96% at pH 5.1 but only 21 to 35% at pH 7.3 (Adaskaveg, Forster, and Wade 2011).

Use appropriate rates. Quantity of active ingredient is important to obtain efficacy. For example, recent work looking at rates of copper products is demonstrating that as metallic copper content increases, copper product efficacy increases up to approximately 0.2 lb metallic copper per 100 gal per acre.

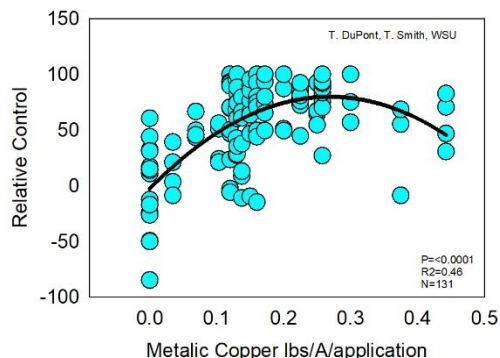


Figure 6 Relative control from coppers WSU trials 2013 to 2017.

Mixtures. A full rate of kasugamycin (100 ppm) with a full rate of oxytetracycline (200 ppm), as well as streptomycin (100 ppm) mixed with a full rate of oxytetracycline (200 ppm) have provided improved efficacy in some trials (Oregon 2015-2018).

Actigard (2oz) plus an antibiotic applied during bloom has improved the efficacy of antibiotics an average of 10% in trials in Washington and Oregon (Smith and Johnson 2011-2014).

Chemical Control Products

Biological Products

When applied to open flowers, these micro-organisms produce colonies on the stigma surfaces and nectary. With biological materials (e.g., Blossom Protect), spray treatments need to be initiated relatively early in the bloom period before high fire blight risk has developed.

Blossom Protect is a combination of two strains of *Aureobasidium pullulans*, a yeast that occurs naturally in Pacific Northwest pome fruit flowers. This organism grows on the nectary and stigmas of treated flowers and competes directing with the fire blight pathogen for the nutritional resource available on these surfaces. Blossom Protect is applied with a companion buffer, Buffer Protect, which reduces the pH of the sprayed suspension and helps the yeast grow faster than the pathogen. In Pacific Northwest trials, Blossom Protect has been the most effective bio-control organism to date (Johnson et al. 2014). If this product is used, it is important to spray every row at least once.

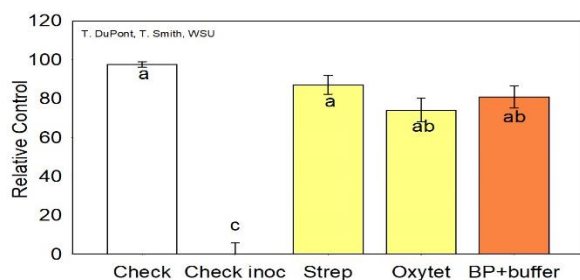


Figure 7 Blossom Protect in WSU Trials 2013, 2014, 2016, 2017.

Antibiotics

Kasugamycin (tradename: Kasumin) is a recently labeled antibiotic that provides **good** levels of control (~80%). All *Erwinia amylovora* strains are currently sensitive to this material but there is an intermediate risk of resistance developing to this antibiotic (Adaskaveg, Forster, and Wade 2011). Kasumin controls streptomycin-resistant strains of *E. amylovora*. Kasumin provides forward control for two to four days prior to rain events (on flowers open when applied) and will be partially effective for blossom blight control if applied within 12 hours after a rain event. Kasumin is not locally systemic like streptomycin. Thus, Kasumin will not penetrate into the nectaries and will not be able to control an infection once the fire blight pathogen reaches the nectaries. Acidifying spray tanks (target 5) is important to reduce antibiotic break down and extend activity.

Oxytetracycline (tradenames: Mycoshield, FireLine) generally provide **good** levels of control in Washington trials and has a low risk of resistance development. Oxytetracycline products should be applied within one day prior to a rain event for best results. Oxytetracycline is considered bacteriostatic (inhibits bacterial growth). Thus, it has to be applied prior to rains where it can prevent growth on stigmas. Oxytetracycline is also sensitive to UV degradation and much of the activity is lost within one to two days after application. Acidifying spray tanks (target 5) is important to reduce antibiotic break down and extend activity.

Streptomycin (tradenames: Agri-Mycin, FireWall): Streptomycin-resistant strains of the fire blight pathogen have been present in Washington orchards since 1975 (Coyier and Covey 1975, Loper et al. 1991). Recent tests have indicated that the proportion of the pathogen population resistant to this antibiotic has dropped, and expected control levels have improved (Forster et al. 2015). This product should only be used in combination with oxytetracycline, and should not be used unless a high to extreme risk infection period is expected. Limit use to once per season. Remaining pathogen colonies in the orchard should be assumed to be streptomycin-resistant.

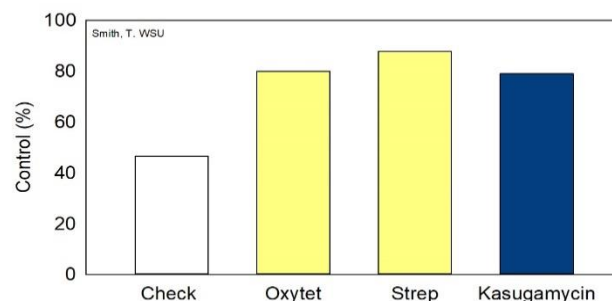


Figure 8 Percent control from antibiotics in WSU Trials 2006, 2009, 2010, 2011.

Coppers

Copper materials vary in the form and amount of metallic copper (the active ingredient). **"Fixed"** (copper hydroxides, copper oxychlorides) -copper products have a longer residual time and are generally used for delayed dormant (green tip) in bearing orchards and summer shoot blight protection in non-bearing (young) orchards. In fixed coppers, most of the copper is insoluble with soluble copper ions released slowly over time. Application of low-pH materials (e.g., Buffer Protect) to trees treated recently with a fixed copper can cause a large release of copper ions and increase the potential for phytotoxicity (Rosenberger 2011). Copper is toxic to plants when a sufficient concentration of ions penetrates tissue. Growers should avoid spray additives such as foliar nutrients and surfactants when applying coppers. Fixed-coppers should not be used with Imidan, Sevin, Thiodan, Captan, or phosphorus acid compounds (Fostphite, Prophyt, Phostrol, Agri-Fos, Alliete) (Shane and Sundin 2011).

Soluble coppers. Newer copper formulations are designed to reduce copper phytotoxicity and fruit russetting potential by introducing far few copper ions to the plant surface and adding safeners that also reduce injury potential. Examples are Cueva (copper octanoate), which is a salt of copper and a fatty acid (copper soap), and Previsto, which is copper ions in a matrix with alginate (polymer from seaweed). Both Cueva and Previsto have shown little phytotoxicity in semi-arid Washington trials but have shown some risk of russetting in wetter areas of Oregon and California. Cueva is compatible in tank-mixes with *Bacillus*-based biopesticides, while Previsto is not due to its high pH.

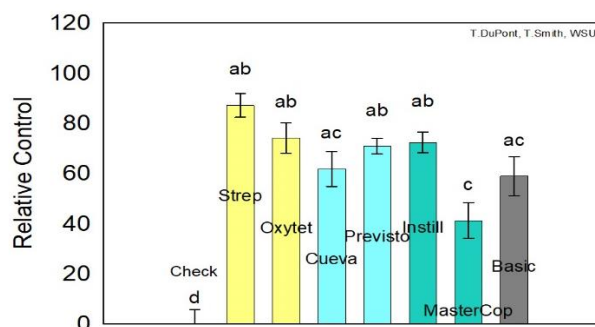


Figure 9 WSU trials 2013, 2014, 2016, 2017. Cueva (copper octanoate) 3-5 qt; Previsto (copper hydroxide) 3-5 qt; Instill (copper sulfate pentahydrate) 1-1.85 qt; Mastercop copper sulfate pentahydrate) 0.25-1.25 qt; Basic coppers Champ, Kocide (0.5 lb), Badge (1.25 pints). Rates/ 100 gal/Acre.

SARs

Acibenzolar-S-methyl (ASM, Actigard 50 WG), is a synthetic inducer of systemic acquired resistance (SAR). Its mode of action is to mimic the plant hormone, salicylic acid, which is responsible for priming the plant's defense system. The level of protection is smaller compared to an antibiotic but it lasts longer, approximately a week (Maxson-Stein et al. 2002).

Biorationals and Biopesticides

Serenade Optimum is an apparently 'fruit safe' material, made by fermenting a strain of *Bacillus subtilis*. The antimicrobial activity of Serenade comes primarily from biochemical compounds produced by the bacterium during fermentation, and not because of the bacterium's colonization of flowers in the orchard.

Apple Materials

Chemical	Rate per Acre	REI	PHI	MOA	Efficacy	Notes
Previsto copper hydroxide	3-4 qt	48 h	none listed		3	Pay attention to drying times and do not combine with acidifying products to reduce fruit finish risks.
Kasumin 2L kasugamycin	64 oz	12 h	90 d		4	Best control when applied less than 48 hrs before wetness event. Control up to 12 hr after wetness event.
DoubleNickel 55 <i>Bacillus amyloliquefaciens</i> strain D747	See label	4 h	0 d		2	See label and space between rows to select the corresponding rate. Efficacy may vary based on disease pressure.
Blossom Protect <i>Aureobasidium pullulans</i>	1.25 lb	4h	none listed		4	30 and 80% bloom. Yeasts need 1-2 days before an infection to colonize the flower before bacteria invade to be effective.
Cueva copper octanoate	4 qt	4 h	0 d		3	Little russet in semi-arid WA trials. Some russet risk in wetter OR. Tank mix compatible with Bacillus-based biopesticides.
FireLine 17WP oxytetracycline	See label	12 h	60 d		4	Best activity within 24 h before wetness event. Check spray tank pH, 5 optimal. 200 ppm: 1.0 lb/100 gal.
Mycoshield oxytetracycline	See Label	12 h	60 d		4	Best activity within 24 h before wetness event. Check spray tank pH, 5 optimal. 200 ppm: 1.0 lb/100 gal.
Actigard 50WG acibenzolar-s-methyl	2 fl oz	12 h	0 d			For bloom applications: Apply 2 oz/A in a tank mix with a fire blight treatment (generally an antibiotic) that is standard in your area. This is generally 2-3 applications between 20% bloom and petal fall depending on the environmental conditions. Do not apply closer than a 7-day interval.
NovaSource Lime Sulfur lime sulfur/calcium polysulfide	2 % v/v	48 h	none listed			Early bloom applications plus oil are antimicrobial. 20 and 70% bloom timings. Reapply biologicals after lime sulfur if used.
Serenade Opti <i>Bacillus subtilis</i> strain QST 713	20 oz	4 h	0 d			Efficacy may vary based on disease pressure.

Pear Materials

Chemical	Rate per Acre	REI	PHI	MOA	Efficacy	Notes
Kasumin 2L kasugamycin	64 fl oz	12 h	90 d		4	Best control when applied less than 48 hrs before wetness event. Control up to 12 hr after wetness event.
Actigard 50WG acibenzolar-s-methyl	2 fl oz	12 h	0 d			For bloom applications: Apply 2 oz/A in a tank mix with a fire blight treatment (generally an antibiotic) that is standard in your area. This is generally 2-3 applications between 20% bloom and petal fall depending on the environmental conditions. Do not apply closer than a 7-day interval.
Blossom Protect <i>Aureobasidium pullulans</i>	1.25 lb	4h	none listed		4	Apply with Buffer Protect. 30 and 80% bloom. Yeasts need 1-2 days before an infection to colonize the flower before bacteria invade to be effective. Russet potential on sensitive varieties in humid conditions.
FireLine 17WP oxytetracycline	1 lb	12 h	60 d		4	Best activity within 24 h before wetness event. Check spray tank pH, 5.5-6.0 optimal. 200 ppm: 1.0 lb/100 gal.
Previsto copper hydroxide	3-4 qt	48 h	none listed		3	Pay attention to drying times and do not combine with acidifying products to reduce fruit finish risks.
Serenade Opti <i>Bacillus subtilis</i> strain QST 713	20 oz	4 h	0 d			
Cueva copper octanoate	4 qt	4 h	0 d		3	Little russet in semi-arid WA trials. Some russet risk in wetter OR. Tank mix compatible with Bacillus-based biopesticides.
Mycoshield oxytetracycline	16 oz	12 h	60 d		4	Best activity within 24 h before wetness event. Check spray tank pH, 5.5-6.0 optimal. 200 ppm: 1.0 lb/100 gal.

Cutting Fire Blight Infections in Season

Cut hard, cut fast

An infected shoot has many millions to billions of pathogen cells. The highest concentration will be near to tip of the branch or infected floral cluster. By cutting a branch we hope to remove many of these cells so that they cannot flow through the tree where they may concentrate in other susceptible tissue and create new infections. Cut AT LEAST 12 to 18 inches below the noticeably infected area into two year or older wood in order to remove the highest concentration of pathogen cells. Young, vigorous or susceptible varieties will require cutting further. Removing infected tissue quickly increases the likelihood of removing more pathogen cells before they invade deeply into the tree. Some recommendations suggest an 'ugly stub cut' where growers make cuts 20-30 cm below visible symptoms into two-year-old or older wood (where resistance is greater due to carbohydrate reserves (Suleman and Steiner 1994) leaving a 10 to 12 cm naked stub. While small cankers will form on many of these cuts, these cankers can be removed during winter pruning (Steiner 2000).

Use of concentrated Actigard during blight clean-up

New research has shown that treatment of trees with the chemical, Acibenzolar-S-methyl (ASM, Actigard 50 WG), may reduce re-occurrence of blight after cutting out infected strikes. Re-occurrence happens when the act of cutting out the disease does not completely remove the pathogen cells that have moved ahead of the expanding canker.

Plants have defense systems. If something stimulates the plant's defense response before the symptoms develop (or re-develop), the plant will be in an active defense mode and will be less affected by disease when it occurs (or re-occurs). Actigard is a compound that has been found to trigger induced resistance. Its mode of action is to mimic the plant hormone, salicylic acid, which is responsible for priming the plant's defense system.

For more than five years, Dr. Ken Johnson of Oregon State University has found that painting a concentrated solution of Actigard on trees after cutting out infection reduced the severity of re-occurring fire blight cankers in pears. For example, he found that without treatment after cutting out fire blight cankers in young Bosc pear trees, the disease came back 50% of the time and began to run through the tree. With Actigard applications, both the proportion of trees in which fire blight came back and the rate of canker expansion was reduced (Johnson and Temple 2016).

During the summer, cut out blight when you see it. Removing a strike can greatly reduce further damage on the tree, especially if you catch the strike early. Apply concentrated Actigard with an up and down motion to a 1/2 meter length of

the central leader or major scaffold near where the fire blight infection was removed. Use the labeled rate of 1 oz/ 1 quart with 1% silicone-based penetrant. One quart will treat approximately 500 cuts.

Additional Resources

Decision Aid System

Visit for the recent model projections of blossom blight risk at your site.

Crop Protection Guide

Crop Protection Guide recommendations are updated on an annual basis.

Organic Fire Blight Management in the Western US

<https://articles.extension.org/pages/74505/organic-fire-blight-management-in-the-western-us>

Dealing with Fire Blight Once it is in the Orchard.

WSU Newsletter article July 2017.

Tips for Using Blossom Protect

WSU Newsletter article April 10, 2017.

Remember last year's infections are this year's risk.

WSU Newsletter article April 2018.

Canker Management

WSU Newsletter article January 2019.

Use pesticides with care. Apply them only to plants, animals, or sites listed on the labels. When mixing and applying pesticides, follow all label precautions to protect yourself and others around you. It is a violation of the law to disregard label directions. If pesticides are spilled on skin or clothing, remove clothing and wash skin thoroughly. Store pesticides in their original containers and keep them out of the reach of children, pets, and livestock.

YOU ARE REQUIRED BY LAW TO FOLLOW THE LABEL. It is a legal document. Always read the label before using any pesticide. You, the grower, are responsible for safe pesticide use. Trade (brand) names are provided for your reference only. No discrimination is intended, and other pesticides with the same active ingredient may be suitable. No endorsement is implied.

Literature Cited

- Adaskaveg, J. E., H. Forster, and M. L. Wade. 2011. "Effectiveness of Kasugamycin Against *Erwinia amylovora* and its Potential Use for Managing Fire Blight of Pear." *Plant Disease* 95 (4):448-454. doi: 10.1094/pdis-09-10-0679.
- Coyier, D. L., and R. P. Covey. 1975. "Tolerance of *Erwinia amylovora* to streptomycin sulfate in Oregon and Washington. ." *Plant Disease* . 59:849-852.
- Elkins, R. B., T. N. Temple, C. A. Shaffer, C. A. Ingels, S. B. Lindow, B. G. Zoller, and K. B. Johnson. 2015. "Evaluation of Dormant-Stage Inoculum Sanitation as a Component of a Fire Blight Management Program for Fresh-Market Bartlett Pear." *Plant Disease* 99 (8):1147-1152. doi: 10.1094/pdis-10-14-1082-re.
- Forster, H., G. C. McGhee, G. W. Sundin, and J. E. Adaskaveg. 2015. "Characterization of Streptomycin Resistance in Isolates of *Erwinia amylovora* in California." *Phytopathology* 105 (10):1302-1310. doi: 10.1094/phyto-03-15-0078-r.
- Johnson, K. B. 2000. "Fire blight of apple and pear." *Plant Health Instructor*. doi: DOI: 10.1094/PHI-I-2000-0726-01.
- Johnson, K. B., V. O. Stockwell, D. M. Burgett, D. Sugar, and J. E. Loper. 1993. "Dispersal of *Erwinia Amylovora* and *Pseudomonas fluorescens* by honey bees from hives to apple and pear blossoms." *Phytopathology* 83 (5):478-484. doi: 10.1094/Phyto-83-478.
- Johnson, K. B., and T. N. Temple. 2016. "Comparison of methods of acibenzolar-S-methyl application for post-infection fire blight suppression in pear and apple." *Plant Disease* (100).
- Johnson, K. B., T. N. Temple, R. B. Elkins, and T. J. Smith. 2014. "Strategy for Non-Antibiotic Fire Blight Control in US-Grown Organic Pome Fruit." In *Xiii International Workshop on Fire Blight*, edited by F. Rezzonico, T. H. M. Smits and E. Holliger, 93-100. Leuven 1: Int Soc Horticultural Science.
- Kleinhempel, H., and M. Nachtigall. 1987. "Disinfection of Pruning Shears for the Prevention of Fire Blight Transmission." *Acta Horticulturae* 217.
- Lecomte, P. 1989. "Risk of Fire blight Infection Associate with Pruning of Pear Trees." *Acta Horticulturae* 273.
- Loper, J. E., M. D. Henkels, R. G. Roberts, G. G. Grove, M. J. Willett, and T. J. . Smith. 1991. "Evaluation of streptomycin, oxytetracycline, and copper resistance in *Erwinia amylovora* isolated from pear orchards in Washington State. ." *Plant Disease* (75):287-290.
- Momol, M. T., J. L. Norelli, D. E. Piccioni, E. A. Momol, H. L. Gustafson, J. N. Cummins, and H. S. Aldwinckle. 1998. "Internal movement of *Erwinia amylovora* through symptomless apple scion tissues into the rootstock." *Plant Disease* 82:646-650.
- Ogawa, J., and H. English. 1991. *Diseases of Temperate Zone Tree Fruit and Nut Crops*. Vol. Publication 3345. Oakland, California: University of California Division of Agriculture and Natural Resources.
- Pattemore, D. E., R. M. Goodwin, H. M. McBrydie, S. M. Hoyte, and J. L. Vanneste. 2014. "Evidence of the role of honey bees (*Apis mellifera*) as vectors of the bacterial plant pathogen *Pseudomonas syringae*." *Australasian Plant Pathology* 43 (5):571-575. doi: 10.1007/s13313-014-0306-7.
- Pusey, P.L., and E.A. Curry. 2004. "Temperature and pomaceous flower age related to colonization by *Erwinia amylovora* and antagonists."
- Rosenberger, D. 2011. Spring Copper Sprays for Fruit Diseases. In *Scaffolds Fruit Journal*. Cornell University.
- Shane, B., and G. W. Sundin. 2011. Copper formulations for fruit crops. edited by Department of Plant Pathology. Michigan State University: MSU Extension.
- Smith, T.J., and P. L. Pusey. 2010. "CougarBlight 2010 ver. 5.0, a Significant Update of the CougarBlight Fire Blight Infection Risk Mode." *Acta Hort* (896):331-338.
- Steiner, P.W. 2000. "Integrated Orchard and Nurseries Management for the Control of Fire Blight." In *Fire Blight: The Disease and its Causative Agent Erwinia Amylovora*, edited by Joel L Vanneste. CABI Publishing.
- Suleman, P., and P.W. Steiner. 1994. "Relationship between sorbitol and solute potential in apple shoots relative to fire blight symptom development after infection by *Erwinia amylovora*." *Phytopathology* 84:1244-1250.
- Teviotdale, B.L. 2011. "Fire Blight."
- Timur Momol, M., and S. Aldwinckle. 2000. "Genetic Diversity and Host Range of *Erwinia amylovora*." In *Fire Blight The Disease and its Causative Agent, Erwinia amylovora*, edited by J. L. Vanneste. New Zealand: CABI.
- Toussaint, V., and V. Pihlon. 2008. "Natural epidemic of fire blight in a newly planted orchard and effect of pruning on disease development." In *Proceedings of the Eleventh International Workshop on Fire Blight*, edited by K. B. Johnson and V. O. Stockwell, 313-+. Leuven 1: Int Soc Horticultural Science.
- van der Zwet, T., and S.W. Beer. 1991. "Fire Blight—Its Nature, Prevention and Control: A Practical Guide to Integrated Disease Management." *U.S. Department of Agriculture, Agriculture Information Bulletin* (631).