



Research

- Diseases- Blackleg. Recent outbreaks in Oregon and Idaho.
 Collaborating with Lindsey du Toit, Jim Davis, and Kurt Schroeder.
- Conducted surveys in 2018





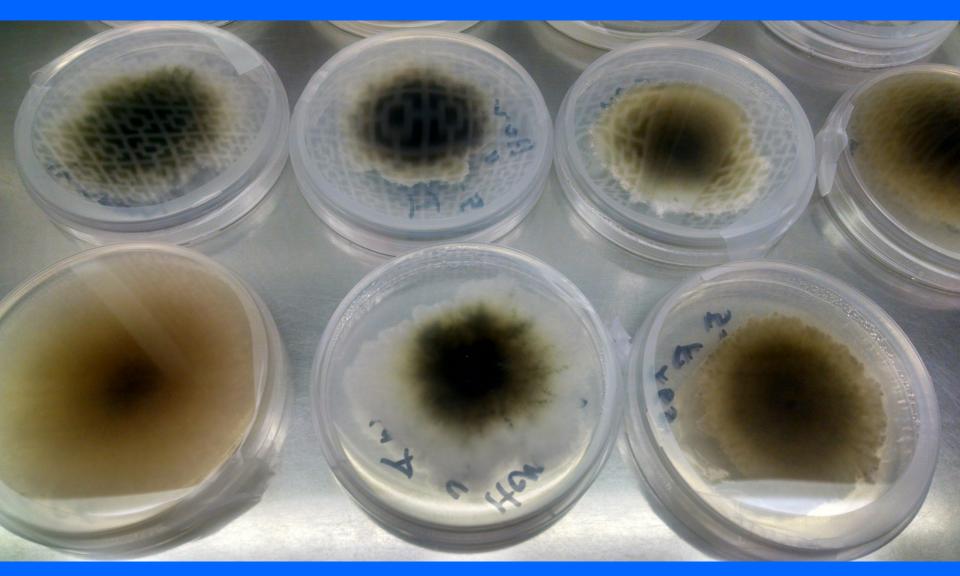


Surveyed 15 locations in Ritzville, Paha, Odessa, Pomeroy, and Genesee.

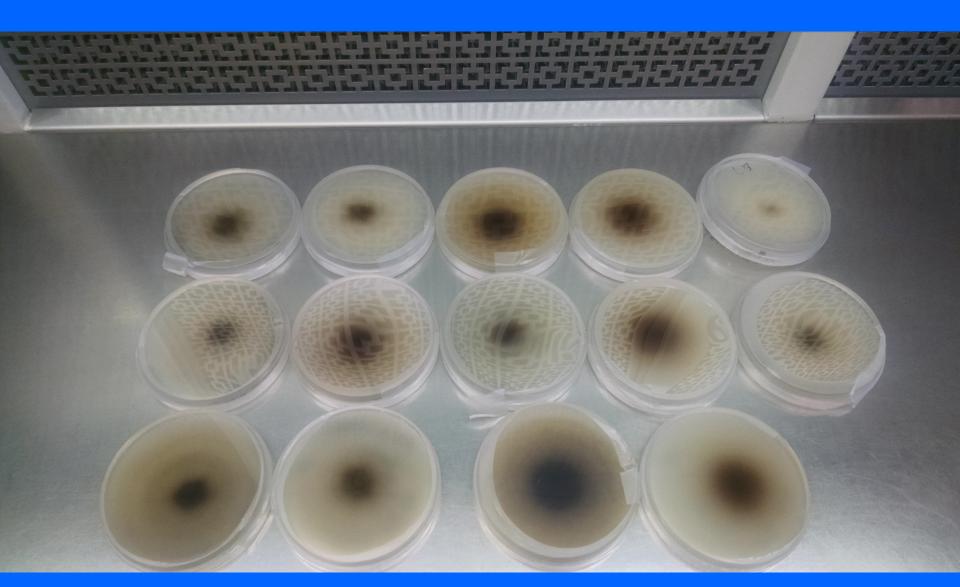
Isolated and plated out and sequenced cultures

Positive finds of *L. maculans* in Odessa, Ritzville, Pomeroy, Genesee and Paha.

Similar results to last yearappears to be established in these areas, although disease severity is low.



Leptospharia maculans



Leptospharia biglobosa

Have a collection of over 100 isolates, to be give to Kurt Schroeder, who has a student starting to work on Leptosphaeria population genetics

Diagnosis of a seed cabbage sample from Whitbey Island- sent by the Washington State Dept. of Agriculture













Was not able to isolate or identify Leptosphaeria, but did find Botrytis



Additional Research with Bill Schillinger

- Spring wheat after winter canola or winter wheat-Reardan (Hal Johnson)
- Consistent yield drag after WC.
- Why? Contrary to most literature.

Additional Research with Bill Schillinger

- Difference in water use?
- Difference in N?
- Herbicides?
- Residue?
- Pathogens? Root lesion nematodes- Winter canola a good host for P. neglectus

Looked at Microbial Communities

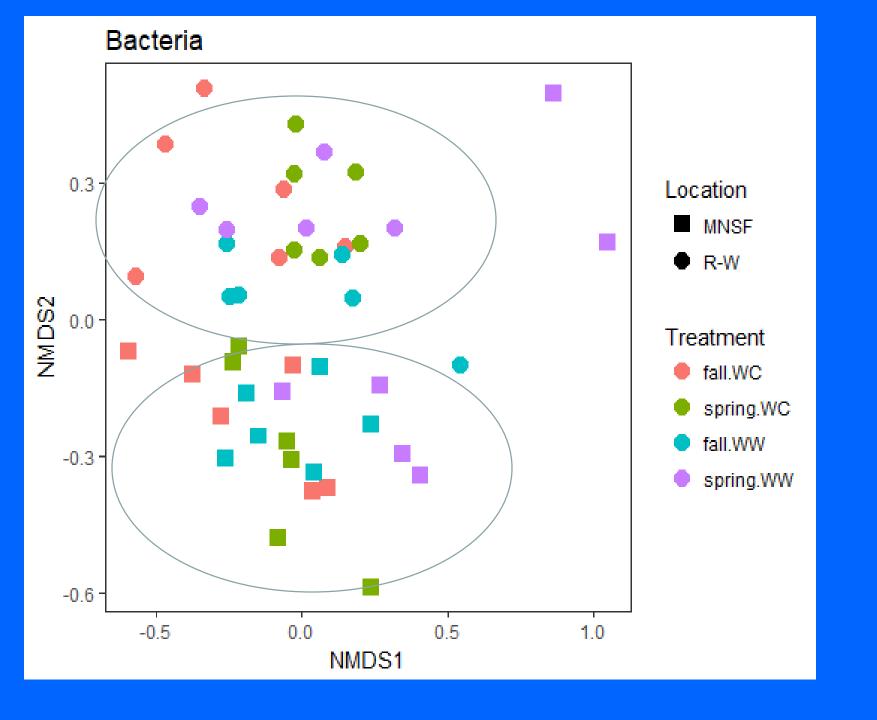
- Sampling by Jeremy Hansen in Ritzville, Washtucna and Mansfield
- Sampled rhizosphere of winter canola and spring wheat
- Did PLFA analysis
- PhD thesis defended in 2018

Looked at Microbial Communities

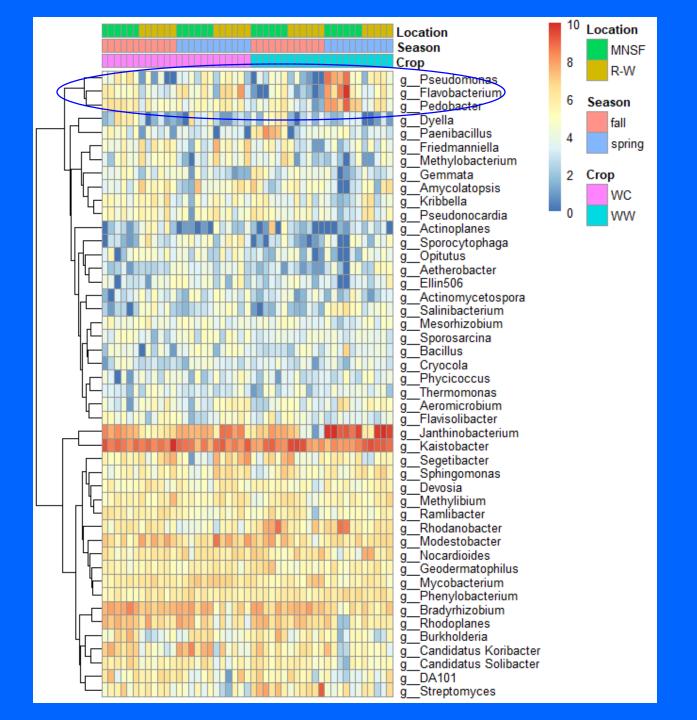
 With Dan Schlatter, did nextgeneration sequencing on DNA from same samples- bacteria and fungi with MiSeq

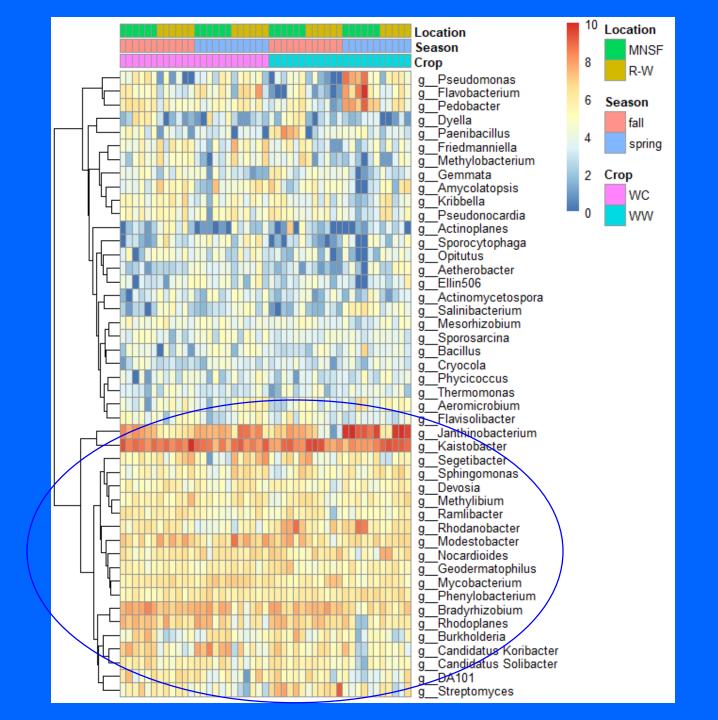
Looked at Microbial Communities

 Question- is there a difference in the rhizosphere communities of winter wheat and winter canola that would explain this yield drag?



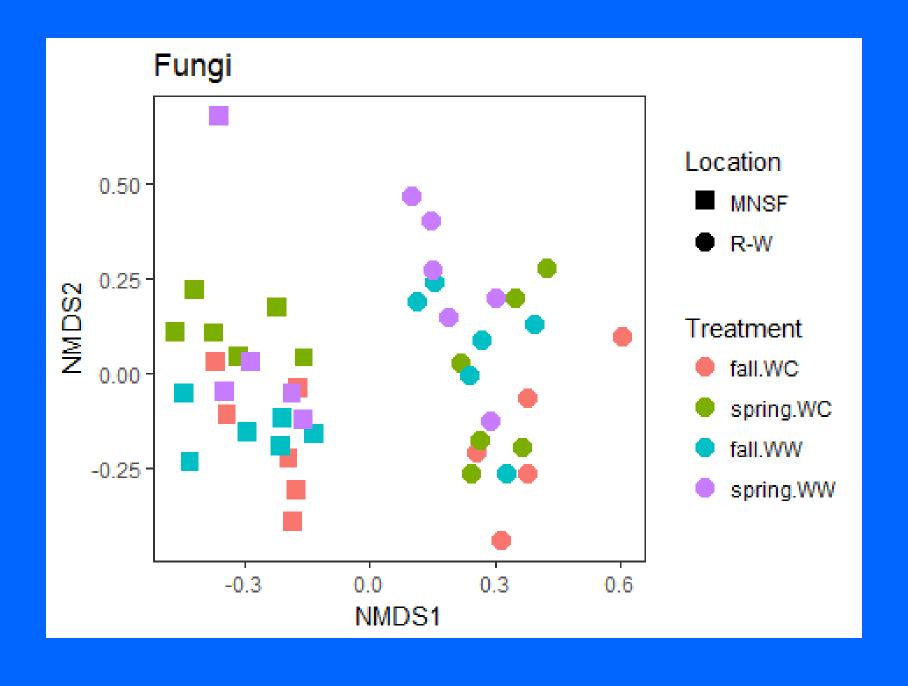
Bacteria	r2	p- value
Dactoria	1 4	value
Season	0.07	0.001
Location	0.12	0.001
Crop	0.04	0.003
Season x Crop	0.04	0.001
Location x Crop	0.05	0.001
Location x Season	0.02	0.09
Location x Season x Crop	0.02	0.11





Was a strong seasonal influence on wheat

- Fall- mostly Actinobacteria and Acidobacteria- oligotrophs, slow growing, survived the hot, dry summer
- Spring- copiotrophs-*Pseudomonas*, Oxalobacteria,
 Flavobacteria, Sphingobacteria



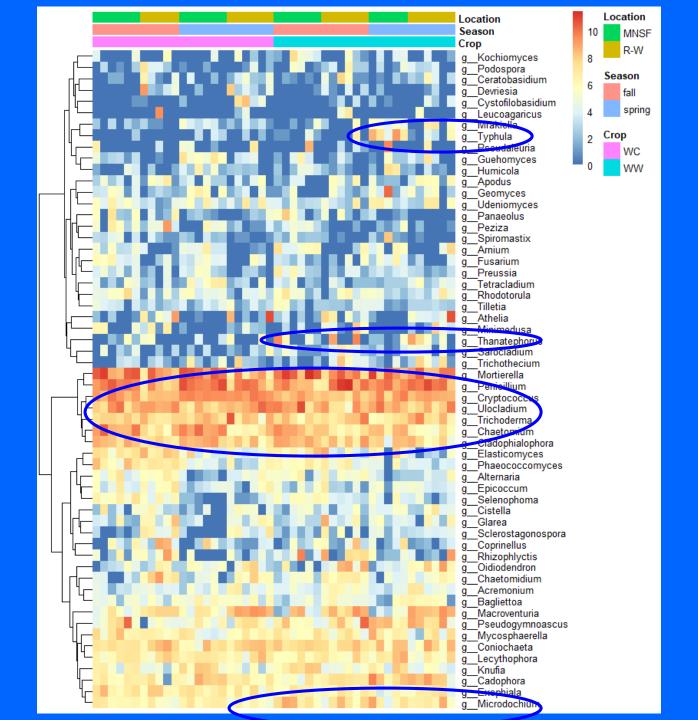
Fungi	r2	p- value
Season	0.05	0.002
Location	0.15	0.001
Crop	0.038	0.008
Season x Crop	0.014	0.7
•		
Location x Crop	0.05	0.001
Location x Season	0.015	0.64
Location x Season x Crop	0.015	0.66

Fungal Communities

- Found 936 OTUs or species
- Large overlap between canola and wheat fungi
- However, many cereal pathogens consistently higher in winter wheat

Fungal Communities

- Thanatephorus cucumerinum (Rhizoctonia solani)
- Ceratobasidium (could be R. cerealis)
- Snow molds- Typhula
- Microdochium



← Wheat

Chytrids higher on canola roots. These are primitive fungi, no report of pathogens on canola roots

Conclusions

- So far, no "smoking gun" of a bacteria or fungus that could be responsible for yield drag
- Canola and wheat share a large group of common bacteria and fungi in the soil

Publications

- Schlatter, D. C., Hansen, J. C., Schillinger, W. F., Sullivan, T. S. and Paulitz, T. C. 2019.
 Common and unique microbial rhizosphere communities in wheat and canola in semiarid Mediterranean environments. Soil Biology and Biochemistry: in revision.
- Hansen, J. C., Schillinger, W. F., Sullivan, T. S. and Paulitz, T. C. 2018. Soil microbial community response with canola introduced into a long-term monoculture wheat rotation. Applied Soil Ecology. 130:185-193.

Publications

- Schillinger, W.F., and T.C. Paulitz. 2018.
 Canola versus wheat rotation effects on subsequent wheat yield. Field Crops Research 223:26-32.
- Hansen, J. C., Schillinger, W. F., Sullivan, T. S. and Paulitz, T. C. 2019. Soil Microbial Biomass and Fungi Reduced with Canola Introduced into Long-Term Monoculture Wheat Rotations. Frontiers in Microbiology: submitted

Products

 Fact Sheet/Grower Handout on Blackleg

Will do a PNW Extension Bulletin in 2019





Blackleg in Canola and other Crucifers What You Need to Know

THE FACTS:

- > The WSDA Crucifer Quarantine now includes all counties of eastern WA
- Blackleg has been confirmed in northeast OR grower fields and at the Pendleton research station
- Lesions have been observed in ID canola and rapeseed fields; the disease has been described as 'common' but not severe
- Blackleg has been confirmed in Garfield Co. based on a few infected leaves of volunteer from the 2015 canola crop

WHEN BUYING SEED:

- > Buy ONLY tested and certified blackleg-free seed
- Look for the green WSDA tag on each seed bag indicating Crucifer Quarantine compliance (including cover crop mixtures)
- > Ask your seed rep for varieties with MR (moderately resistant) or R (resistant) blackleg rating
- > Apply seed treatment (most companies already do but double check that)

AFTER EMERGENCE:

- > Scout fields for any lesions on leaves and/or cankers on stems (see back for photos)
- > Continue to monitor fields throughout the growing season

SCOUTING PROTOCOL to avoid spreading blackleg

- Wear rubber boots
- > When finished scouting/sampling a field, scrape and wash off any soil adhering to boots
- Spray boots with 70% alcohol (isopropyl alcohol works well)
- Remove boots and wear clean shoes until reaching the next field

IF BLACKLEG IS OBSERVED (current crop or past crop residue)

- Place fresh leaves and/or stems in a ziploc bag. If they are wet, blot them dry on a paper towel
- Mail (preferably overnight) or deliver samples to the WSU Plant Diagnostic Clinic, UI or OSU Plant Pathology departments (see contact info on next page)
- Follow recommendations for applying fungicide ONLY if blackleg is confirmed and at or above threshold levels

HARVEST and TRANSPORTATION

- > Make sure combine is set properly to reduce as much seed loss as possible
- > Tarp trucks and seal up rear gates and belly dumps before delivery

OTHER KEY RECOMMENDATIONS

- > Control Brassica/crucifer volunteers and weeds in fields and field borders
- > Rotate canola and other brassicas; grow no more than once every 3 years on the same field
- > Learn how to identify blackleg symptoms; be vigilant in scouting fields







WSU contacts:

Tim Paulitz USDA-ARS

Room 363 Johnson Hall Pullman, WA 99164-6430 509-335-7077 | 509-592-6401

Rachel Bomberger

WSU Plant Pest Diagnostic Clinic 316 Johnson Hall Pullman, WA 99164-6430 509-335-0619

University of Idaho contact:

Kurt Schroeder

PSES Dept. Ag Sci. Bldg. Rm 242 Moscow, ID 83844 208-885-5020

Oregon State University contact:

Don Wysocki

509-278-4396 | 541-969-2014

Send samples to:

Robert Cating

OSU Extension Plant Pathology Lab, HAREC

2121 South 1st Street

Hermiston, OR 97838





Extension

 Presented talks on diseases of oilseeds at workshop in Colfax in Feb. 2018

Future Work

- PNW extension publication on black leg
- New project with Bill Schillinger on canola yield drag at Ritzville.
 Observing a similar phenomenon as Reardan
- Opportunity to do microbial community work and sequencing

